



**vivantis**  
Nucleic Acid Extraction Kit HandBook

***GF-1***

PLANT DNA EXTRACTION  
USER GUIDE (Version 1.2)

Catalog No.

SAMPLE: 5 preps

GF-PT-050: 50 preps

GF-PT-100: 100 preps

High Yield and Purity

Fast and Easy purification

Reliable and Reproducible

Eluted DNA ready for use in downstream applications

No toxic or organic-based extraction required

## Introduction

The **GF-1 Plant DNA Extraction Kit** is designed for rapid and efficient purification of genomic DNA from a variety of plant tissues without the need for precipitation or organic extraction. This kit uses a specially-treated glass filter membrane fixed into a column to efficiently bind DNA in the presence of high salt. The kit applies the principle of a mini-column spin technology and the use of optimized buffers to ensure that only DNA is isolated while proteins and other impurities, additives, preservatives are removed during the subsequent washing steps. High purity genomic DNA is then eluted in water or low salt buffers has a  $A_{260/280}$  ratio between 1.7 and 1.9, making it ready to use in many routine molecular biology applications such as restriction enzyme digestion, Southern blotting, DNA fingerprinting, PCR and other manipulations.

## Kit component

Product Catalog No.	5 Preps SAMPLE	50 Preps GF-PT-050	100 Preps GF-PT-100
<b>Components</b>			
GF-1 columns	5	50	100
Collection tubes	5	50	100
Plant Tissue Lysis Buffer ( <b>Buffer PL</b> )	2ml	18ml	36ml
Plant Genomic Binding Buffer ( <b>Buffer PB</b> )	4ml	35ml	70ml
Wash Buffer (concentrate)*	3ml	24ml	2 x 24ml
Elution Buffer	1ml	10ml	20ml
Proteinase K*	0.11ml	1.05ml	2 x 1.05ml
Handbook	1	1	1

\* Please refer to **Reconstitution of Solutions** and **Storage and Stability** before using this kit.

The **GF-1 Plant DNA Extraction Kit** is available as 50 and 100 purifications per kit.

The reagents and materials provided with the kit are for research purposes only.

Note: The **GF-1 Plant DNA Extraction Kit** is optimized to isolate up to 20µg of DNA from 10-30mg of plant tissue. Plant tissue samples vary in the number of cells depending on age, type of tissue and origin. When processing samples, and do not use more than the recommended starting material as excessive number of cells will overload the column. This would result in reduced yield and purity. We recommend weighing the tissue samples before starting to ensure optimum yield and purity is obtained.

### **Additional Materials to be Supplied by User**

Absolute Ethanol (>95%)

RNase A (DNase-free) (20mg/ml)

### **Reconstitution of Solutions**

The bottle labeled **Wash Buffer** contains concentrated buffer which must be diluted with absolute ethanol (>95%) before use.

For **SAMPLE (5 preps)**,

Add **7ml** of absolute ethanol into the bottle labeled **Wash Buffer**.

For **GF-PT-050 (50 preps)**,

Add **56ml** of absolute ethanol into the bottle labeled **Wash Buffer**.

For **GF-PT-100 (100 preps)**,

Add **56ml** of absolute ethanol into one of the bottles labeled **Wash Buffer**.

Add **56ml** of absolute ethanol into the other bottle labeled **Wash Buffer** only prior to use.

Store **Wash Buffer** at room temperature with bottle capped tight after use.

### **Storage and Stability**

- Store solutions at 20°C - 30°C.
- Store **Proteinase K** at -20°C.
- Kit components are guaranteed to be stable for 12 months from the date of manufacture. **Buffer PB** may exhibit salt precipitation due to cold temperature. If this occurs, simply warm the bottle at 55°C - 65°C with occasional mixing until precipitate is completely dissolved.

**Chemical Hazard**

**Buffer PB** contains guanidine salts which can be harmful when in contact with skin or swallowed. Always wear gloves and practice standard safety precautions. Do NOT disinfect guanidine or extraction waste in solutions containing bleach or any other form of acid. To clean any items contaminated with the reagent, simply soak in detergent and water to remove all traces of guanidine before cleaning with bleach or acidic solution

## Procedures

### Reminder

- All steps are to be carried out at room temperature unless stated otherwise.
- **Wash Buffer** (concentrate) has to be diluted with absolute ethanol before use.  
Please refer to **Reconstitution of Solutions**.
- If precipitation forms in **Buffer PB**, incubate at 55°C - 65°C with occasional mixing until completely dissolved.

Pre-set waterbath to 65°C.

Pre-heat **Elution Buffer** at 65°C (optional).

#### 1. Homogenization

Cut 10 - 30mg of tissue sample into small pieces with a clean scalpel. Freeze sample in liquid nitrogen and grind into a fine powder with a mortar and pestle.

*The tissue sample should be ground into fine powder to ensure complete lysis in the next step.*

#### 2. Tissue Lysis

Add 280µl of **Buffer PL** to the ground sample. Mix thoroughly by vortexing the tube for 30 sec to obtain a homogeneous solution. Add 20µl of **Proteinase K** and mix thoroughly by inverting tube. Incubate at 65°C for 1 - 2 hrs (or overnight if tissue mixture does not appear clear) in a shaking waterbath or mix several times during incubation to ensure thorough digestion of the sample.

*If tissue sample was not reduced to small pieces, homogenize sample in buffer with multiple strokes using a tube pestle. Solubilization of tissue sample varies between different tissue types. If insoluble materials still remain, extend incubation time or increase the amount of Proteinase K to ensure complete lysis.*

### 3. Centrifugation

Centrifuge at 14,000 - 16,000 x g for 5 min to precipitate any insoluble/undigested materials. Transfer the supernatant containing the DNA into a clean microcentrifuge tube.

*Extend centrifugation time if solids are not completely spun down.*

#### **Optional: Removal of RNA**

If RNA-free DNA is required, add 20µl of RNase A (DNase-free, 20mg/ml). Mix and incubate at 37°C for 5 min.

*Residual RNA fragments will be removed during column washing.*

### 4. Homogenization

Add 2 volumes (~600µl without RNase A treatment, ~640µl with RNase A treatment) of **Buffer PB** and mix thoroughly until a homogeneous solution is obtained by inverting tube several times. Incubate for 10 min at 65°C.

*Precipitation may occur due to high DNA content. Lysate should be clear upon mixing and incubation at 65°C.*

### 5. Addition of ethanol

Add 200µl of absolute ethanol. Mix immediately and thoroughly.

*Mix immediately to prevent any uneven precipitation of nucleic acid due to high local ethanol concentrations.*

### 6. Loading to column

#### ***Attention! Tips for Higher Yield***

*In order to obtain maximum yield, we strongly recommend users to fix the orientation of the column during centrifugation at all times. We recommend users to place the column which has a triangle mark on the edge, at a fixed position during centrifugation.*

Transfer the sample into a column (max. 900µl) assembled in a clean collection tube (provided). Centrifuge at 10,000 x g for 1 min. Discard flow through.

Repeat for the remaining sample from step 6.

*If column clogs, add 200µl Buffer PB into column and centrifuge as above.*

## 7. **Column washing**

Wash the column with 750µl **Wash Buffer** and centrifuge at 10,000 x g for 1 min. Discard flow through. Repeat washing if color stains from sample remains on the column membrane.

*Ensure that ethanol has been added into the Wash Buffer before use (refer to Reconstitution of Solutions).*

## 8. **Column drying**

Centrifuge the column at 10,000 x g for 1 min to remove residual ethanol.

*This step has to be carried out to remove all traces of ethanol completely as residual ethanol can affect the quality of DNA and may subsequently inhibit enzymatic reactions.*

## 9. **DNA elution**

Place the column into a clean microcentrifuge tube. Add 50 - 100µl of preheated **Elution Buffer**, TE buffer or sterile water directly onto column membrane and stand for 2 min.

Centrifuge at 10,000x g for 1 min to elute DNA. Store DNA at 4°C or -20°C.

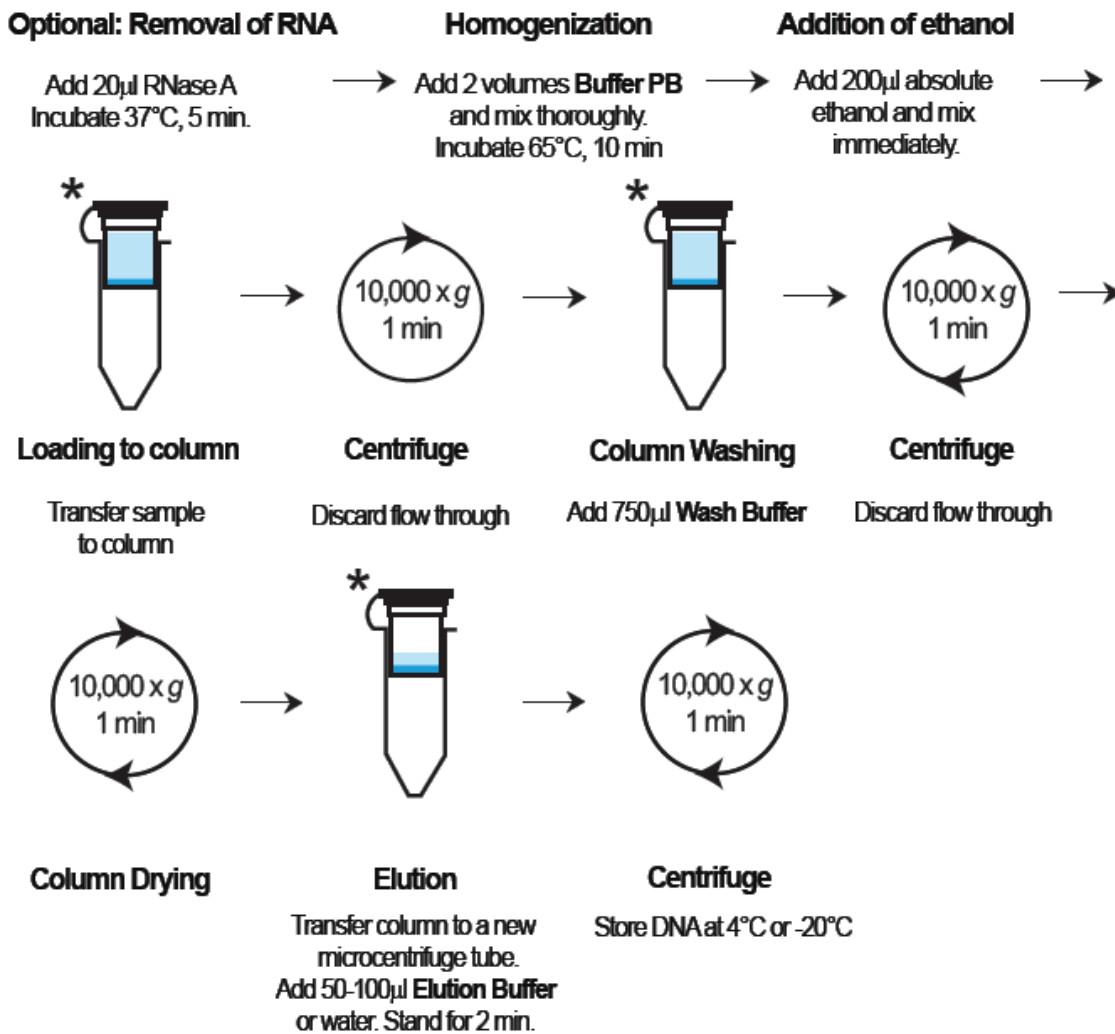
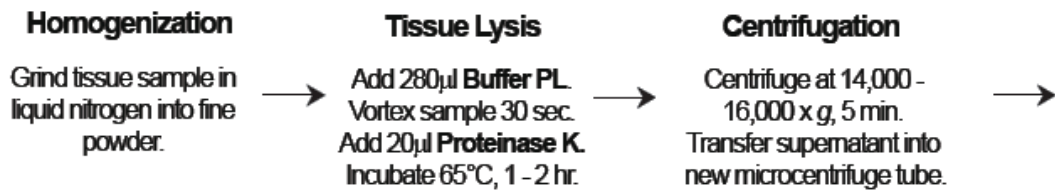
*Ensure that the Elution Buffer is dispensed directly onto the center of the membrane for complete elution. TE Buffer can also elute DNA although EDTA may inhibit subsequent enzymatic reactions. If water is used for eluting DNA, maximum elution efficiency is achieved between pH7.0 and 8.5. Store DNA at -20°C as DNA may degrade in the absence of a buffering agent.*

## Troubleshooting

Please note that by not adhering to the recommended protocols, unsatisfactory results related to yield and quality of DNA may occur. If problems arise, please refer to the following:

<b>Problem</b>	<b>Possibility</b>	<b>Suggestions</b>
<b>Low DNA yield</b>	<i>Sample not thoroughly homogenized</i>	<i>Ensure that tissues are completely homogenized in <b>Buffer PL</b></i>
	<i>Samples not fresh or not properly stored</i>	<i>For long term storage of tissues, keep at -70°C.</i>
	<i>Sample not lysed completely</i>	<i>Ensure that tissues are completely homogenized in <b>Buffer PL</b> and mix sample frequently during incubation in absence of a waterbath shaker.</i>
	<i>Low elution efficiency</i>	<i>Pre-heat <b>Elution Buffer</b> to 65°C-70°C before eluting DNA.</i>
	<i>Column clogged</i>	<i>Refer to Problems under 'Column clogged'.</i>
	<i>Column not placed at fixed orientation during centrifugation</i>	<i>Place the column which has a triangle mark on the edge, at a fixed position during centrifugation at all times.</i>
<b>Low purity</b>	<i>Incomplete protein denaturation</i>	<i>Extend incubation time until lysate clears.</i>
	<i>RNA contamination</i>	<i>Add RNase A to the sample as indicated in the protocol. Ensure that RNase A used has not been repeatedly frozen and thawed. If necessary prepare a fresh stock</i>
<b>No DNA eluted</b>	<i>Inappropriate elution buffer</i>	<i>Ensure that the <b>Elution Buffer</b> used is a low salt buffer or water with a pH range of 7.0-8.5</i>

<b>Problem</b>	<b>Possibility</b>	<b>Suggestions</b>
<b>Column clogged</b>	<i>Overloading of column/ starting material too high</i>	<i>Do not use more than 30mg of sample material. If any undigested material remain, spin to remove tissue lysate and transfer supernatant into a new microcentrifuge tube.</i>
	<i>Sample not thoroughly homogenized</i>	<i>Vortex sample in <b>Buffer PL</b> prior t addition of <b>ProteinaseK</b></i>
<b>DNA degradation/ smearing</b>	<i>DNA sheared during purification</i>	<i>After the addition of <b>Buffer PL</b> and <b>Proteinase K</b>, avoid vigorous mixing and pipetting. Mix gently by inverting tube.</i>
	<i>Nuclease contamination</i>	<i>Use sterilized glassware, plasticware and wear gloves.  Ensure that the tissue is completely homogenized in <b>Buffer PL</b> and <b>Proteinase K</b></i>
<b>Poor performance of eluted DNA in downstream applications</b>	<i>Eluted DNA contains traces of ethanol</i>	<i>Ensure that the Column drying step is carried out prior to elution.</i>



**\* We recommend users to place the column which has a triangle mark on the edge, at a fixed position during centrifugation.**